



Integrated Study of Food borne Antimicrobial Resistance (AMR) in Kenya

Patrick Otto

Animal Health Officer (Veterinary Public Health)

FAO Headquarters, Rome

Patrick.otto@fao.org



Outline

1. Introduction
2. Study scope and Objectives
3. Study design and Sampling
4. Methodology
5. Provisional Results
6. Follow-up activities, outputs/outcomes





FAO mandate

“Food security exists when all people, at all times, have physical, social and economic access to sufficient, **safe and nutritious** food” [FAO World Food Summit, 1996]

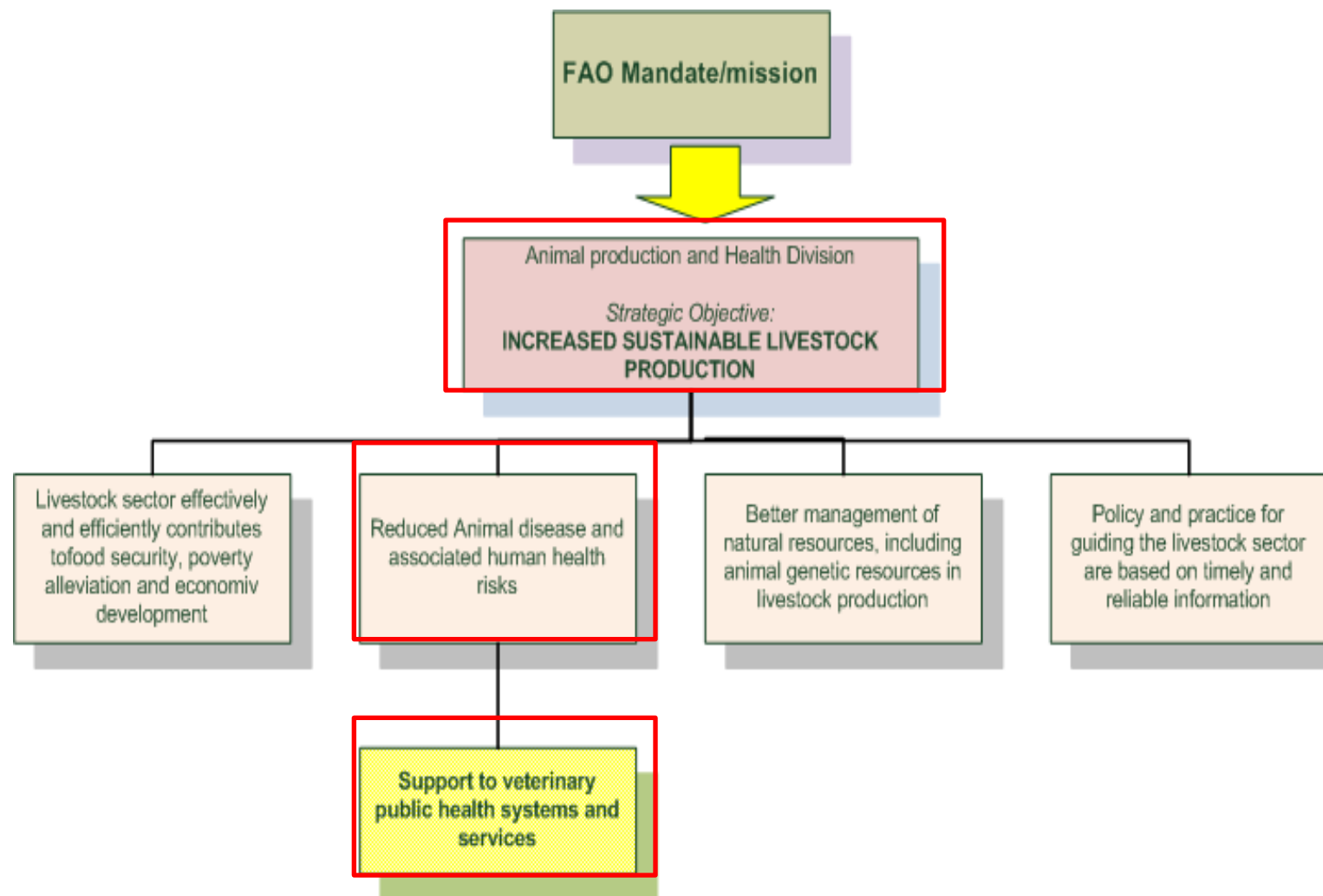


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Food Safety is key to health and nutrition = the ultimate goals of Food Security



Strategic Framework and Context





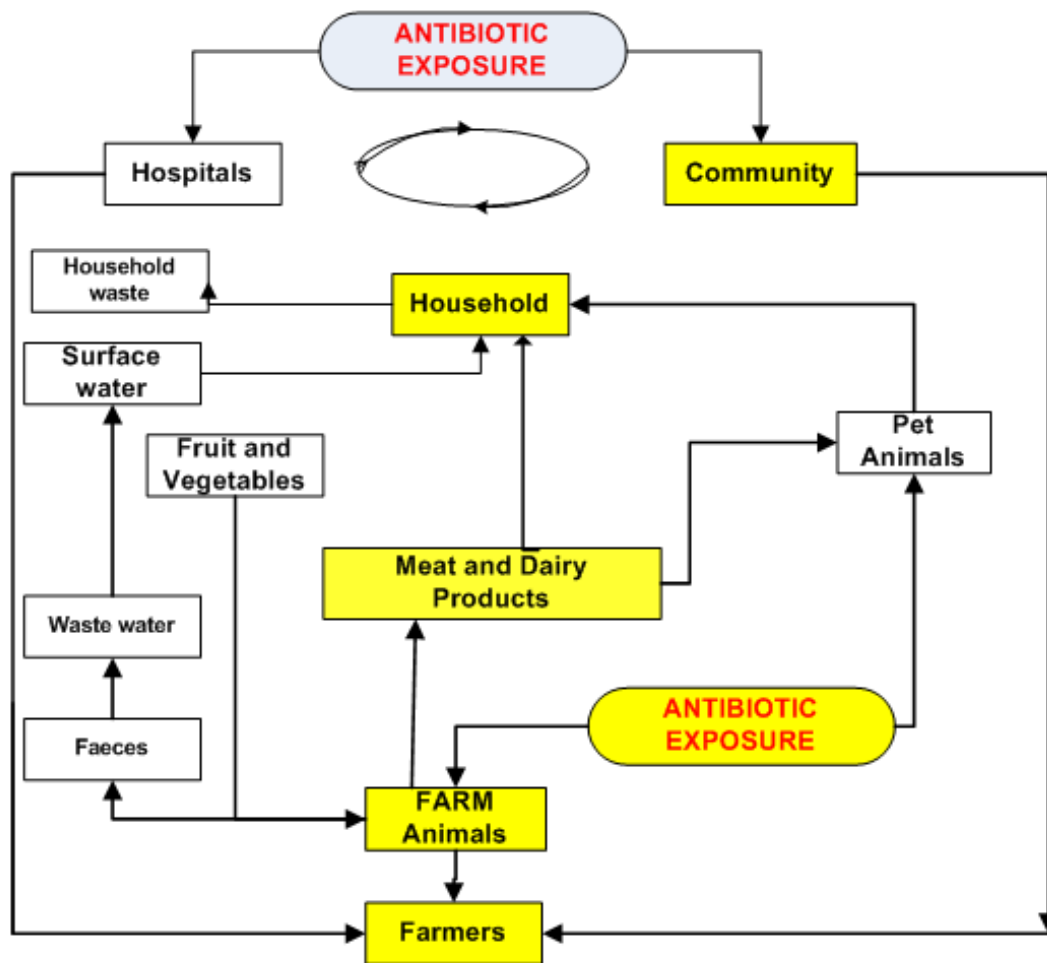
FAO AMR objectives

- Promote prudent and responsible use of antimicrobial drugs: support livelihoods of livestock owners and economies; and minimize human health risks;
- support national/regional capacities to manage AMR risks – at all stages of the food chain;
- Collaboration with international partners, esp. WHO and OIE in development of tools/guidelines and contribute to standards setting functions.



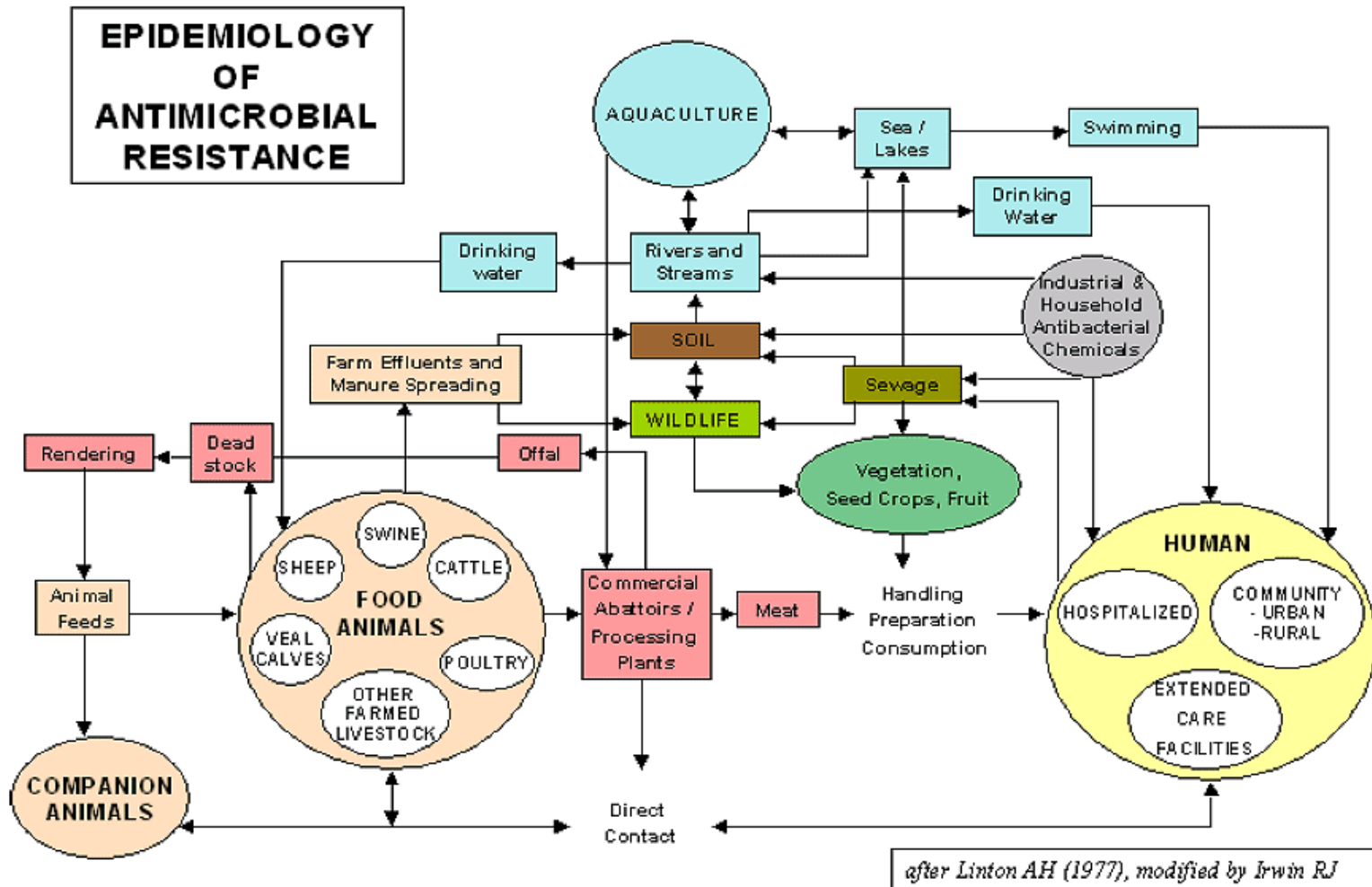


AMR pathways: *exposure and spread*



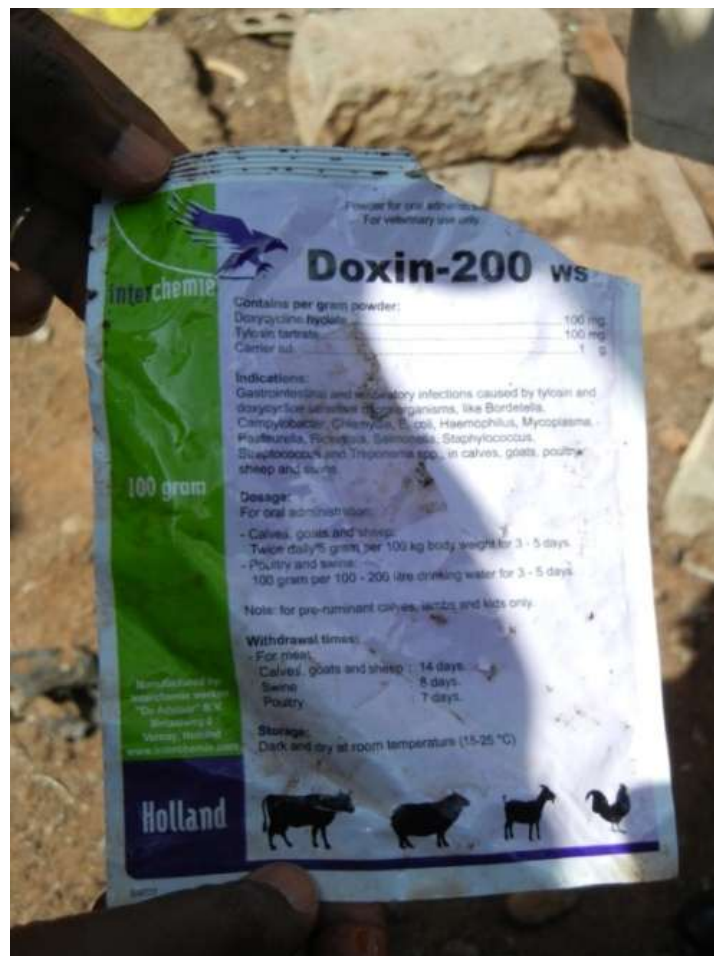


Integrated Program for Antimicrobial Resistance Surveillance (CIPARS)





Usage of veterinary antimicrobials





Overall Objective of Kenya study

To undertake a baseline integrated prevalence study of food pathogen contamination and antimicrobial resistance in farm animals, abattoirs and retail meat outlets - as well as from human clinical specimens in selected sites in Kenya.



- FAO /WHO Collaboration
- Implementing partner: *Kenya Medical Research Institute (KEMRI)*



Specific Objectives

1. To determine the prevalence of *Salmonella* spp, *Campylobacter* spp, *E. coli* and *Enterococcus* spp in food animals, carcasses and retail meats as well as from human clinical specimens in selected regions in Kenya - ongoing
2. To determine the antimicrobial susceptibility patterns of these foodborne pathogens to commonly available classes of antimicrobials - ongoing
3. To investigate *in-vitro* transferability of antibiotic resistance determinants in *E. coli* and *Enterococcus* spp (indicator organisms) isolated from animals and meat products – pending
4. Beef/Poultry/(Pig) Value Chain Assessments
- value chain assessment, review of national food safety information/data and analysis of food safety institutional framework - completed.



Study sites



● Study sites

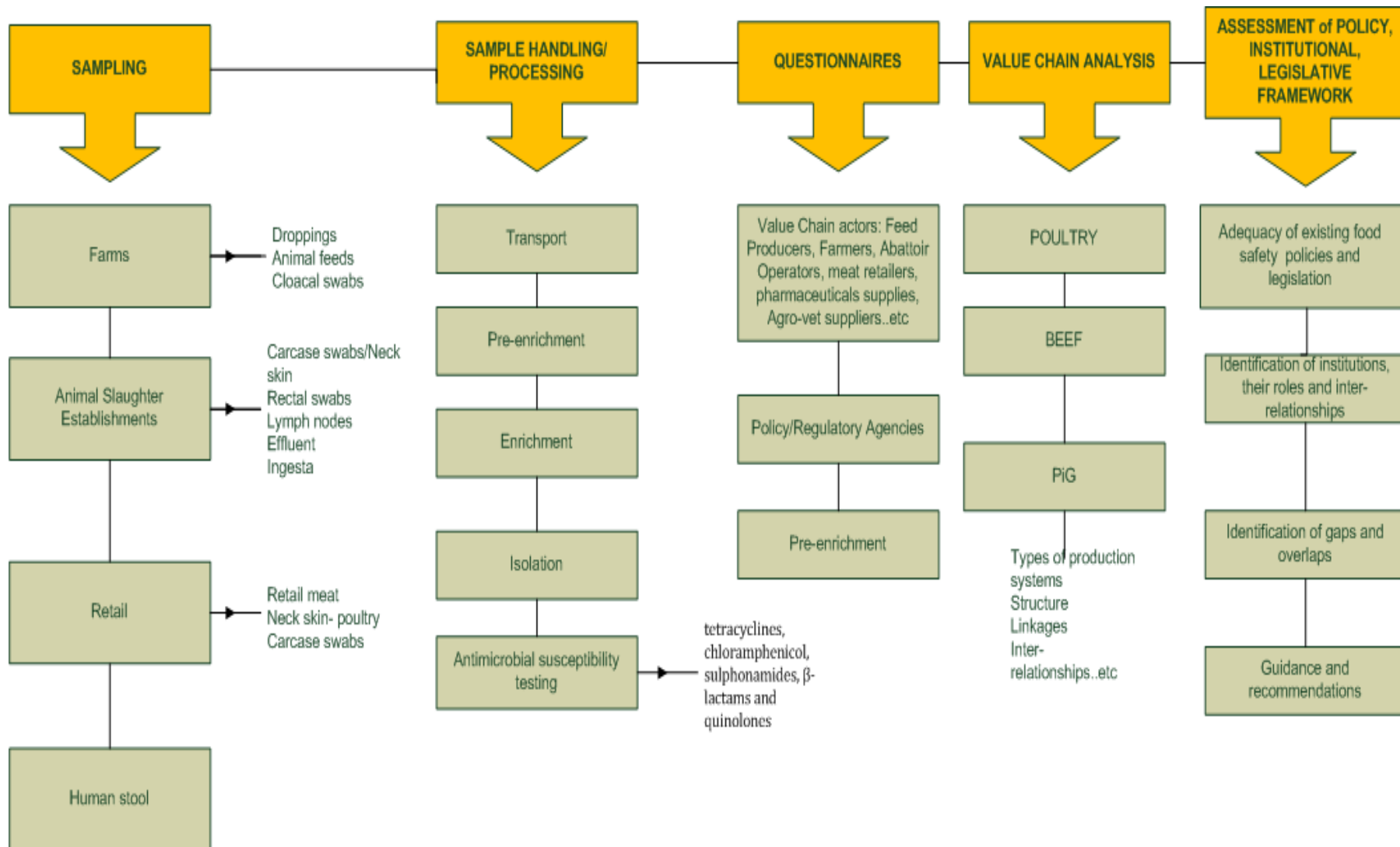
1. **Nairobi and Thika County** – farms, main public abattoirs run by the City and municipal authorities and meat outlets served by these abattoirs
2. **Nyanza**: abattoirs, farms and outlets in Kisumu, Maseno and Kakamega
3. **Coast**: slaughter houses, farms and outlets in Mombasa, Kwale and Malindi

Differences in:

- size,
- rural/urban,
- production systems,
- management and hygiene standards



Study Design and Methods





Sampling

- Feed Producers
 - *Animal Feeds*
- Farms
 - *Animal feeds*
 - *Cloacal/rectal swabs*
 - *Droppings*
- Animal Slaughter Establishments
 - *Carcase swabs/Neck skin*
 - *Ingesta/intestinal contents*
 - *Lymph nodes*
 - *Effluent*
- Retail
- Clinical specimens

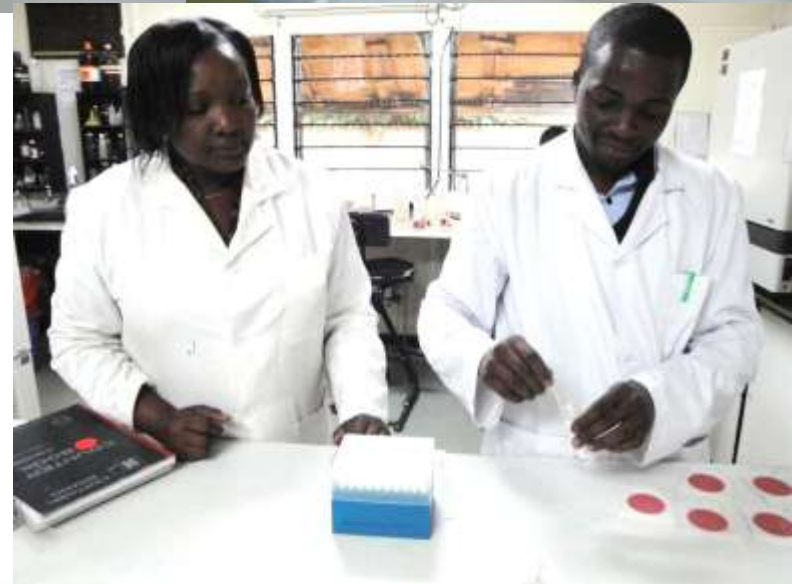
Experts Panel advice on sampling and study methodology





Laboratory procedures: *isolation & enumeration*

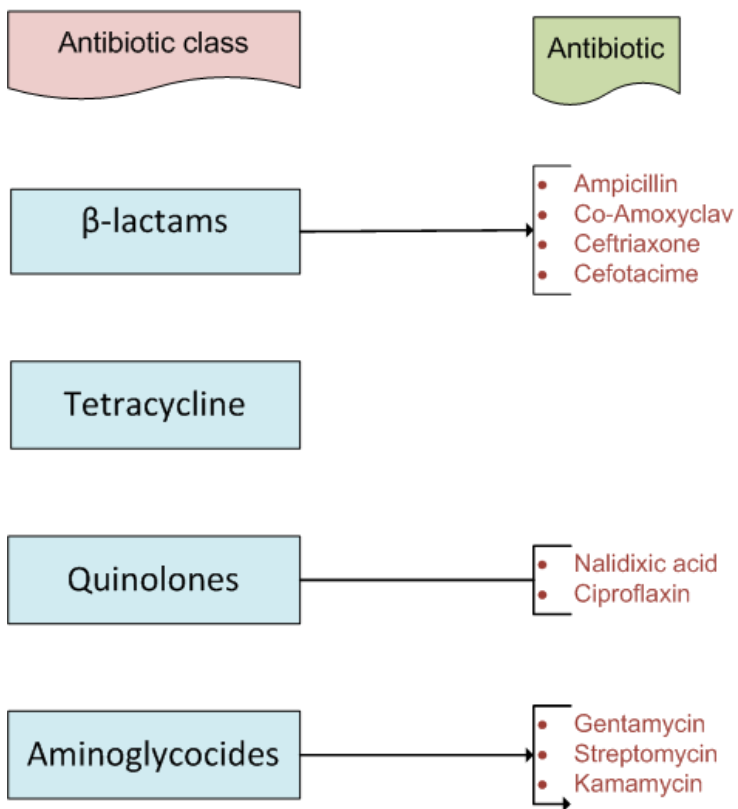
- Isolation: Enrichment, selective enrichment, biochemical tests and Gram stain for identification:
 - *Salmonella* spp
 - *E. coli*
 - *Enterococcus* spp
 - *Campylobacter* spp
- Enumeration of total *E. coli* and coliform counts by 3M Petrifilm plate method





Antibiotic susceptibility Tests

Antibiotic Susceptibility Tests – Kirby-Bauer Disk diffusion Technique (Bauer et al. 1966)



Campylobacter spp. Isolates – all tested for susceptibility to Erythromycin, tetracycline and ciproflaxin (agar dilution method)

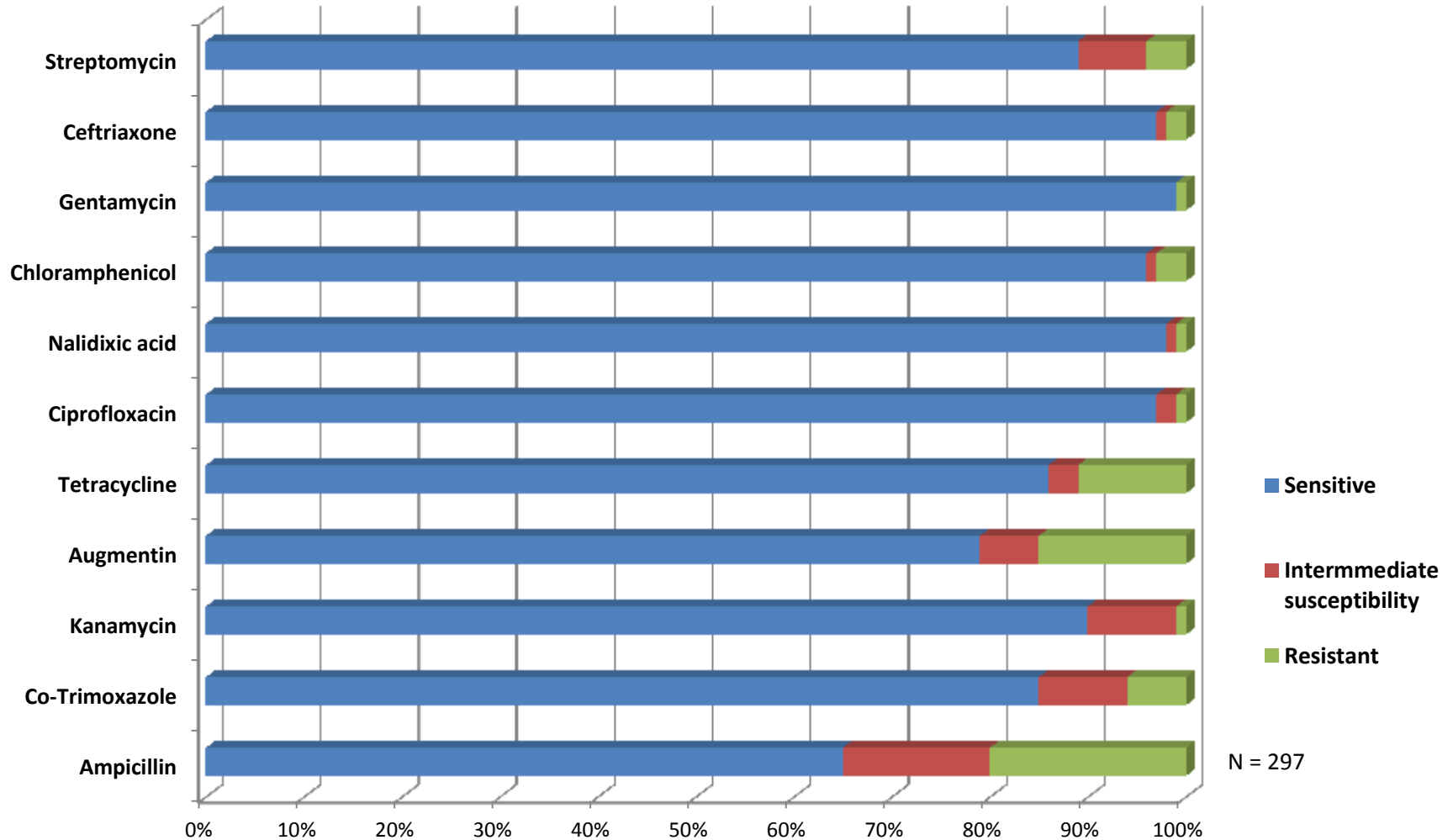


Total bacterial isolations per each type of specimen

Type of animal sampled	Total no. of specimen sampled	Type and total no. of bacteria isolated
Cattle	1492	1053 E. coli, 12 Salmonella spp., 33 Enterococcus spp., 6 Citrobacter spp., 9 Providencia spp., 2 Kluyvera spp., 5 Enterobacter spp., 1 Tatumella spp., 1 Pseudomonas spp., 1 Budvicia spp., 28 Campylobacter spp., 1 Vibrio metschiikonii, 1 Klebsiella spp., 3 Shigella spp., 2 Yersinia spp., 5 Proteus spp., 7 Aeromonas spp.
Pig	69	48 E. coli, 1 Citrobacter spp., 2 Klebsiella spp.
Chicken	1792	1254 E. coli, 18 Salmonella spp., 2 Citrobacter spp., 2 Klebsiella spp., 10 Proteus spp., 1 Morganella spp., 1 Pseudomonas spp., 16 Enterococcus spp., 172 Campylobacter spp.
Effluent	181	139 E. coli, 12 Salmonella spp., 1 Citrobacter spp., 4 Enterococcus spp., 2 Campylobacter spp., 1 Tatumella spp., 1 Proteus spp., 2 Providencia spp.
Chicken feeds	75	64 E. coli, 7 Enterococcus spp.

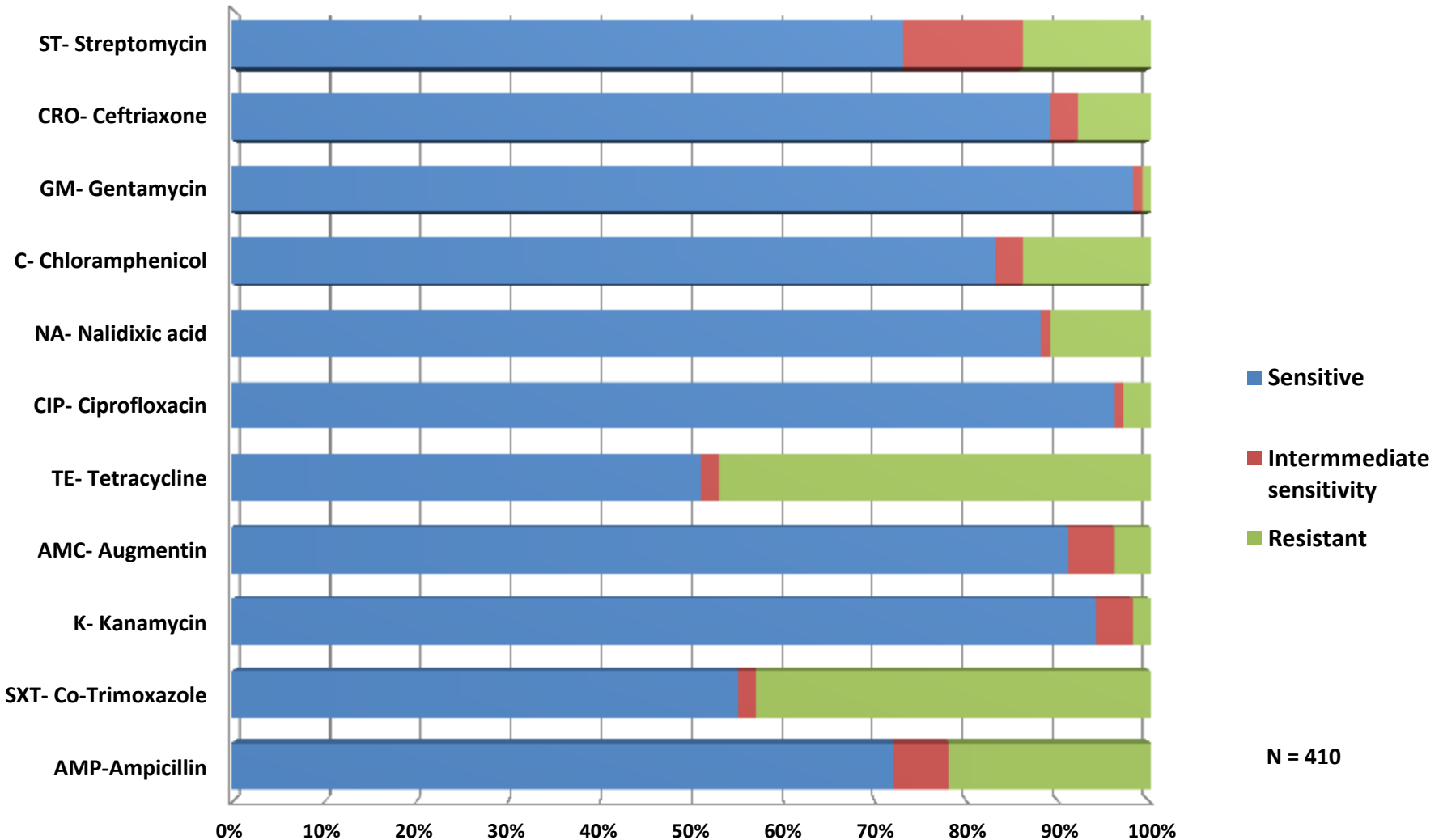


Beef Isolates: Antibiotic susceptibility patterns for *E.coli*





Poultry Isolates: Antibiotic susceptibility patterns for *E. coli* isolates

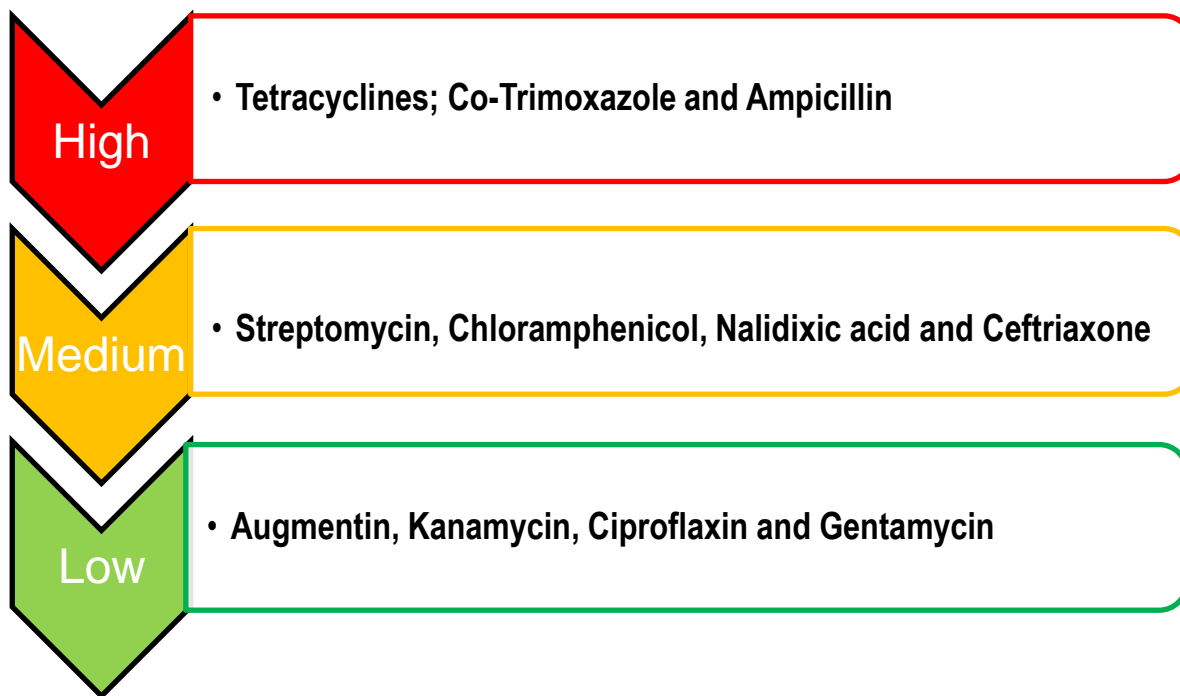




Prioritization of interventions

Codex Guideline for Risk Analysis of Food borne AMR (CAC/GL-2011)

- *Risk Assessment*
- *Risk management*
- *Surveillance of use of AM agents and AMR organisms and AMR determinants*
- *AMR Risk Communication*





Follow-up activities and outputs/outcomes

1. Regional Technical Meeting (August 2011): to review preliminary results and discuss follow-up actions in context of regional harmonization;
2. Support development and implementation of national/regional programmes on surveillance of use of antimicrobial agents, and AMR;
3. Other collaborative projects - *replication of initiative in other developing countries*;
4. Development guidance and support implementation:
 - *Policy*
 - *Prudent use of antimicrobials in animal production*
 - *Risk-based hygiene controls in abattoirs*



Thank You

Patrick.otto@fao.org